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Detailed insights into pan-European population structure and inbreeding in wild and hatchery Pacific oyster (*Crassostrea gigas*) populations revealed by genome-wide SNP data

Running head: population structure and inbreeding in Pacific oyster populations

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Abstract

Cultivated bivalves are hugely important not only because of their economic value, but also due to their impacts on natural ecosystems. The Pacific oyster (*Crassostrea gigas*) is the world's most heavily cultivated shellfish species and has been introduced to all continents except Antarctica for aquaculture. We therefore used a medium density single nucleotide polymorphism (SNP) array to investigate the genetic structure of this species in Europe, where it was introduced during the 1960s and has since become a prolific invader of coastal ecosystems across the continent. We analyzed 21,499 polymorphic SNPs in 232 individuals from 23 localities spanning a latitudinal cline from Portugal to Norway and including the source populations of Japan and Canada. We confirmed the results of previous studies by finding clear support for a southern and a northern group, with the former being indistinguishable from the source populations indicating the absence of a pronounced founder effect. We furthermore conducted a large-scale comparison of wild and hatchery populations to reveal substantial genetic differences including significantly higher levels of inbreeding in some but not all of the hatchery populations. These findings were confirmed by a smaller but representative SNP dataset generated using restriction site associated DNA sequencing. We therefore conclude that genomic approaches can generate increasingly detailed insights into the genetics of invasive populations, while also providing a novel window on how hatchery propagation may influence inbreeding, with important economic and management implications.

Keywords:

Pacific oyster, *Crassostrea gigas*, single nucleotide polymorphism (SNP), high density genotyping array, restriction site associated DNA (RAD) sequencing, aquaculture, genetic structure, inbreeding

Introduction

Oysters are among the most economically important aquaculture species, with worldwide annual production exceeding 600,000 tonnes (FAO, <http://www.fao.org>). In particular, the Pacific cupped oyster (*Crassostrea gigas*), which is native to the Pacific coast of eastern Asia, was introduced into many countries worldwide for commercial cultivation. Starting in the 1960s, *C. gigas* was introduced into Europe to support oyster farming after severe declines of the two previously cultivated oyster species—the Portuguese oyster (*C. angulata*) and the flat oyster (*Ostrea edulis*, Grizel & Héral 1991, Nehring 1999, Wolff & Reise 2002). Large quantities of seed as well as adult oysters were brought to France and the Netherlands from the Miyagi prefecture in Japan and from British Columbia in Canada, where *C. gigas* was also introduced from Japan in the 1920s (Quayle 1988), and became quickly established in the wild. Concurrently, several small importations of less than a hundred individuals at a time also took place into the United Kingdom for hatchery propagation (Walne & Heln 1979).

Subsequently, Pacific oysters produced in UK hatcheries were farmed in the German Wadden Sea (Reise 1998) as well as in Denmark (Nehring 2006), while oysters produced in French farms were transferred to various locations in the Mediterranean Sea (Grizel & Héral 1991, Šegvić-Bubić et al. 2016) including southern Portugal, where hybridization with *C. angulata* is known to occur (Huvet et al. 2004, Batista et al. 2017). More recently, *C. gigas* also reached the southern coasts of Sweden and Norway (Troost 2010), where it arrived as a consequence of both natural dispersal from Denmark and human-mediated translocation from British hatcheries (d'Auriac et al. 2017). Consequently, Pacific oysters have become widespread across the Atlantic and Mediterranean coasts of Europe, where they are responsible for major changes to coastal ecosystems (Troost 2010) and are considered an invasive species (Goulletquer et al. 2002).

Several studies have used genetic markers such as mitochondrial DNA and microsatellites to investigate the population structure of the Pacific oyster in Europe, mainly with a view towards understanding the history of invasion (Meistertzheim et al. 2013, Rohfritsch et al. 2013, Lallias et al. 2015, d'Auriac et al. 2017, Faust et al. 2017) as well as interrelationships between wild and hatchery populations (Moehler et al. 2011, Kochmann et al. 2012, Lallias et al. 2015). Many of these studies uncovered evidence for two distinct genetic clusters: one in southern Europe (subsequently referred to as the 'southern group') that includes populations from the Mediterranean, Spain, France, the Netherlands and the south-western coast of England, and one in northern Europe (subsequently referred to as the 'northern group') that consists of the remaining British, German and Scandinavian populations (Moehler et al. 2011, Meistertzheim et al. 2013, Rohfritsch et al. 2013, Lallias et al. 2015). Furthermore, no genetic differences were found between the southern group and the source populations of Japan and British Columbia, suggesting that the original mass introduction may not have resulted in a founder effect (Rohfritsch et al. 2013). Additionally, the northern group was found to have lower genetic diversity, suggesting that it probably arose locally in Europe and more specifically in the UK as a consequence of repeated small introduction events that may have acted as bottlenecks due to hatchery propagation followed by genetic drift (Faust et al. 2017, Lallias et al. 2015).

Although previous studies have provided important insights into the population structure of Pacific oysters in Europe, many focused on local scales and, even though Rohfritsch et al. (2013), Lallias et al. (2015) and Faust et al. (2017) sampled extensively along the western Atlantic seaboard, there is still a need for more comprehensive studies encompassing the full latitudinal range of the species in Europe and including hatcheries from both Britain and France. Furthermore, classical approaches like mitochondrial sequencing and microsatellite genotyping have limited power to detect population structure, especially over fine geographic scales where genetic differences may be too subtle to be captured with a handful of markers (Vendrami et al. 2017). By contrast, new genomic approaches capable of genotyping tens of thousands of single nucleotide polymorphisms (SNPs) have been

proven to have far greater power to resolve genetic differences among populations (Morin et al. 2004, Rašić et al. 2014) and therefore allow more in-depth studies of population genetic structure.

One of the most commonly used approaches for generating large SNP datasets for non-model organisms is to use genotyping by sequencing methods such as restriction site associated DNA (RAD) sequencing (Baird et al. 2008), which allows concurrent SNP identification and genotyping via high throughput sequencing of flanking regions of restriction enzyme digestion sites dispersed throughout the genome. These methods have democratized the study of population genomics but are not without their disadvantages (da Fonseca et al. 2016) such as the need for extensive bioinformatic processing, high rates of missing data and the issue of uncertainty in genotype calling, which can affect downstream analyses (Shafer et al. 2017). A convenient alternative where available is therefore to use a medium or high-density SNP array, in which the probe sequences of many tens or hundreds of thousands of SNPs are ‘printed’ onto a slide against which the genomic DNA is hybridized. SNP arrays typically generate very high quality data with relatively few missing genotypes, but they also have some downsides. Arguably the most important of these is ascertainment bias, which occurs when not all of the genetic diversity present in a population can be captured by the array due to the use of a limited pool of individuals in the original SNP discovery phase (Lachance & Tishkoff 2013).

Another drawback of small panels of nuclear markers like microsatellites is that their sampling variance is usually too large to accurately quantify variation in inbreeding (Balloux et al. 2004). This may be relevant to aquaculture because inbreeding has been shown to have detrimental effects on a variety of commercially important traits, such as harvest body size and larval growth, in several species, including turbot (Lyu et al. 2018), Pacific white shrimp (Moss et al. 2007), coho salmon (Gallardo et al. 2004) and Pacific abalone (Deng et al. 2005) as well as in flat oysters (Lallias et al. 2010) and Pacific oysters (Launey & Hedgecock 2001, Evans et al. 2004, Plough & Hedgecock 2011). Hence, inbreeding depression could conceivably be a problem if aquaculture practices, such as the use of restricted numbers of parents as broodstock and / or the crossing of related individuals, lead to increased inbreeding in cultured populations (Norris et al. 1999, Taris et al. 2007).

Given the low power of microsatellites to quantify inbreeding, the method of choice until recently has been to derive individual inbreeding coefficients (f) from deep pedigrees (Pemberton 2008). However, pedigrees can be costly and time consuming to construct and may also be unworkable for many aquaculture species due to their high fecundity and broadcast spawning life-histories. Fortunately, recent simulation (Kardos et al. 2015, Wang 2016) and empirical (Hoffman et al. 2014, Huisman et al. 2016) studies suggest that inbreeding can now be directly and accurately quantified from genomic data, with around ten thousand or more SNPs being preferable under most circumstances even to a high quality pedigree. Consequently, the increasing availability of SNP arrays for non-model species provides an exciting new opportunity to elucidate how different aquaculture practices influence inbreeding, as well as to identify suitable sources of individuals for use as broodstock to establish effective management and breeding protocols.

Recently, Gutierrez et al. (2017) developed a medium density combined species SNP array for Pacific and flat oysters (*Ostrea edulis*). Whole genome sequencing of pooled genomic samples from eight European *C. gigas* populations led to the discovery of 1.2 million putative SNPs, of which 40,625 were printed on the array and 27,697 were validated as being polymorphic and of high quality. This array represents an important resource for selective breeding programs as well as more generally for population genetic studies of oysters. We therefore used it to investigate population genetic structure and inbreeding in wild and hatchery European *C. gigas* populations. Specifically, we genotyped 192 individuals from 13 populations spanning a European latitudinal cline from Portugal in the south to Norway in the north. We then incorporated existing data from Gutierrez et al. (2017) to generate a combined dataset of 273 individuals sampled from 23 populations, of which just over half were wild. Our results may be useful for designing exchanges among hatcheries, identifying potential sources of broodstock, and for the elaboration of other effective management strategies aimed at minimizing inbreeding within hatchery propagated Pacific oyster populations. Consequently, we believe this study provides important information for breeding programs as well as a baseline for future studies.

Materials and methods

Sample collection and DNA extraction

Pacific oyster samples were collected between November 2014 and March 2016 from 12 different sites along the Atlantic seaboard of mainland Europe as well as from one location in the Mediterranean (Table 1 and Figure 1). The populations from Scotland (SCO) and Wales (WAL) were from hatcheries while the remaining 11 populations were wild. Specimens from Portugal (POR) originated from an area where hybridization between *C. angulata* and *C. gigas* is known to take place (Batista et al. 2017, Huvet et al. 2004) and could therefore represent *C. gigas* samples introgressed with *C. angulata*. For comparison, we also included samples from the Miyagi Prefecture in Japan (JAP) and from British Columbia in Canada (CAN).

DNA extraction and SNP genotyping

Adductor muscle tissue was taken from each adult oyster and stored in 95% ethanol at -20 °C. Whole genomic DNA was then extracted following an adapted phenol-chloroform protocol (Sambrook et al. 1989) and sent to Edinburgh Genomics for genotyping at 40,625 SNPs on the custom Affymetrix SNP Array (Gutierrez et al. 2017). Out of a total of 204 DNA extracts, 192 (94%) passed quality checks and were therefore selected for genotyping using the protocol described by Gutierrez et al. (2017).

Incorporation of existing data

We also incorporated data into our study from 81 oysters that were previously genotyped on the same array (Gutierrez et al. 2017). These samples were initially included in the discovery panel used to develop and validate the SNP array and came from eight hatchery populations, three from the UK (MAL, SES and GUE) and five from France (FH1–4 and IFR, Table 1 and Figure 1).

After the inclusion of these additional samples, our dataset consisted of (i) six wild populations from the southern group (red circles in Figure 1); (ii) five hatchery populations from the southern group (orange circles in Figure 1); (iii) four wild populations from the northern group (blue circles in Figure 1); (iv) five hatchery populations from the northern group (purple circles in Figure 1); (v) the source

populations of Japan and Canada (yellow circles in Figure 1); and (vi) a single population from Portugal, where hybridization between *C. angulata* and *C. gigas* is known to occur (green circle in Figure 1).

SNP calling

We imported raw output data for all 273 samples into the Axiom Analysis Suite (version 3.1, Affymetrix) for quality control and genotype calling. All thresholds for quality assessment were set to the values recommended in the Affymetrix best practice workflow (Supplementary Table 1) and allowed for the categorization of each SNP into one of six possible classes: (i) “polymorphic high resolution” where the SNP passed all quality controls; (ii) “no minor homozygote” where the SNP passed quality checks but no homozygotes for the minor allele were found; (iii) “off-target variant” where, in addition to the heterozygote and the two alternative homozygotes, a fourth genotype cluster was also observed; (iv) “monomorphic high resolution” where the SNP passed quality checks but was uninformative; (v) “call rate below threshold” where the genotype call rate was below the specified threshold of 97%; and (vi) “other” where the SNP failed to pass any other quality threshold.

Following Affymetrix recommendations, SNPs from the first two categories were retained for further analysis, in addition to a subset of SNPs from the third category that were selected after applying the “off-target caller” tool that allows for off-target variant recalibration. The resulting dataset was then filtered to retain only SNPs genotyped in at least 90% of individuals and only samples with less than 5% missing data. Finally, the software PLINK (version 1.9, Purcell et al. 2007) was used to prune out linked loci using an r^2 threshold of 0.5. The final dataset therefore comprised 232 individuals genotyped at 21,499 polymorphic, unlinked SNPs.

RAD sequencing

To provide a comparison with the SNP array data, we also RAD sequenced a representative subset of 40 individuals from eight populations (Supplementary table 2). Specifically, we included the source population of Japan (JAP), the potential hybrid population from Portugal (POR), two geographically distant wild populations from the southern group (SPA and NE2), two wild populations from the

northern group (DEN and NOR), the Mediterranean population from Italy (ITA) and a hatchery population from Scotland (SCO). Only one farmed population could be included because DNA from the other hatcheries was either not of high enough quality to pass thresholds for library construction, or it was not available due to the sample having been genotyped as part of a previous study (Gutierrez et al. 2017). Whole genomic DNA was shipped to the Beijing Genomics Institute (BGI) for library preparation and sequencing. The libraries were constructed using the restriction enzyme PstI and sequenced on an Illumina X Ten platform to generate a total of 869,113,776 100 bp paired-end sequence reads. Already demultiplexed sequence data were received from BGI and further sequence quality assessment was performed using the software FastQC (<http://www.bioinformatics.babraham.ac.uk/projects/fastqc/>). We then conducted a *de novo* assembly of the data and called genotypes using the Stacks 2.1 pipeline (Catchen et al. 2013). Values for the three main parameters $-m$, $-M$ and $-n$ were chosen following the optimization procedure described by Rochette & Catchen (2017). Briefly, $-m$ was set to three, and increasing values for $-M$ and $-n$ were tested. The combination of these parameters for which the number of polymorphic loci present in at least 80% of the individuals reached a plateau was defined as optimal. Two different strategies were employed: $-n$ was either set as equal to $-M$ or one unit greater, to account for the potential presence of fixed *C. angulata* polymorphisms (Paris et al. 2017). The combination yielding the highest plateau ($m = 3$, $M = 5$ and $n = 6$; Supplementary Figure 1) was selected for analyzing the entire dataset, from which PCR duplicates were then removed. The raw genotypes were subsequently quality filtered to retain only biallelic SNPs with both genotype quality and depth of coverage greater than 10 using VCFTools (Danecek et al. 2011). Subsequently, all SNPs and individuals with genotyping rates below 10% were removed and only variants with minor allele frequency (MAF) greater than 0.05 were retained. Finally, the software PLINK was employed to prune out linked loci using an r^2 threshold of 0.2.

Analysis of population structure

Three complimentary approaches were used to characterize the strength and pattern of population structure. First, we calculated pairwise F_{st} values among populations and determined their statistical significance based on 1,000 permutations of the dataset using Arlequin version 3.5.2.2 (Excoffier & Lischer 2010). We then performed an Analysis of Molecular Variance (AMOVA) employing the R package ‘poppr’ version 2.8.0 (Kamvar et al. 2014, Kamvar et al. 2015) to evaluate the proportion of genomic variation explained by different levels of hierarchical population structure. Specifically, we started by dividing our data into four regions corresponding to the northern group, southern group, Portugal and source populations. Each was then sub-divided into wild and hatchery samples, which were further split based on the population of origin. Variation among samples within each population and within individuals was also evaluated. A randomization test with 1,000 repetitions was used to determine statistical significance. Subsequently, we used Mantel tests to evaluate the significance of isolation-by-distance patterns for the full dataset as well as for the wild and hatchery populations separately for the northern and southern groups. For this analysis, geographic distances between populations were calculated as shortest coastline distances using “FreeMapTools (<https://www.freemaptools.com/measure-distance.htm>)”. In the case of locations not connected by land, the shortest sailing distance between coasts was used. Second, we used the R package Adegenet version 2.1.1 (Jombart 2008, Jombart & Ahmed 2011) to conduct a principal component analysis (PCA) of the SNP dataset. Third, we utilized the software package fineRADstructure (Malinsky et al. 2018) to infer population structure using a model-based Bayesian clustering approach that groups together individuals with high levels of shared coancestry. A “coancestry matrix”, defined as a summary of nearest neighbor haplotype relationships, is required as input and was generated using the ‘RADpainter’ module of fineRADstructure. We used the default parameters of 100,000 Markov chain Monte Carlo (MCMC) iterations with a burn-in of 100,000 iterations and sampling occurring every 1,000 iterations. A tree was then constructed with 10,000 hill-climbing iterations and the results were visualized using the scripts FINERADSTRUCTUREPLOT.R and FINESTRUCTURELIBRARY.R, which are available via <http://cichlid.gurdon.cam.ac.uk/fineRADstructure.html>.

Genomic inbreeding coefficients

We calculated \hat{F}_I , a genomic inbreeding estimator based on the variance of additive genotype values (Yang *et al.* 2011), for each individual in our dataset based on the SNP data. To test for an association between levels of relatedness and inbreeding, we calculated mean pairwise relatedness among individuals within populations from the SNP data using GCTA (Yang *et al.* 2011) and correlated this with mean \hat{F}_I values. Genomic inbreeding coefficients were compared between populations and groups using a Kruskal-Wallis test followed by *post hoc* pairwise Mann-Whitney tests, whose *p*-values were adjusted according to Benjamini & Hochberg (1995), to formally test for significant pairwise comparisons. As variation in inbreeding causes heterozygosity to be correlated across loci, we also estimated the extent of identity disequilibrium (ID, Weir & Cockerham 1973) by calculating the two-locus heterozygosity disequilibrium, g_2 (David *et al.* 2007) within the R package inbreedR (Stoffel *et al.* 2016). The same package was also used to calculate the 95% confidence interval of g_2 by bootstrapping the data 1000 times over individuals, as described by Stoffel *et al.* (2016).

Results

To provide detailed insights into the pan-European population structure of *C. gigas* and facilitate comparisons between wild and hatchery populations, we analyzed medium-density SNP array data for a total of 273 individuals sampled from 23 populations. Data from 192 individuals were newly generated while the remaining data were incorporated from Gutierrez *et al.* (2017). Sampling sites were putatively assigned to either the northern or the southern group on the basis of previous genetic studies (Rohfritsch *et al.* 2013, Lallias *et al.* 2015). Application of the filtering criteria described in the Materials and methods resulted in the exclusion of 17,411 SNPs that did not meet Affimetrix recommendations and of additional 1,715 SNPs due to low genotyping rate or linkage disequilibrium. On average 10 individuals were genotyped for each location, and the final dataset consisted of 232 samples (see Table 1 for a breakdown by population) genotyped at 21,499 SNPs.

Population genetic structure

To investigate broad-scale patterns of genetic differentiation, we used AMOVA to quantify the proportion of genomic variation attributable to each of five hierarchical levels of population substructure. As expected, over 95% of the total variation was partitioned within individuals. The remaining variance was mainly partitioned among the northern and southern groups, the source populations and Portugal ($\Phi=0.017$, $p=0.024$, Table 2), between wild and hatchery populations ($\Phi=0.004$, $p=0.005$, Table 2) and among populations ($\Phi=0.02$, $p=0.001$, Table 2). Furthermore, the majority of pairwise F_{st} values between populations were highly significant, even after correction for multiple tests (Supplementary table 3), although a significant isolation-by-distance pattern was only detected among wild populations belonging to the southern group (Mantel's $r = 0.971$; $p = 0.022$).

To evaluate population structure at the individual level, we performed a principal component analysis (PCA). Consistent with the AMOVA, a number of clear differences were apparent. First, the northern and southern groups clearly separated apart from one another, as did oysters from Portugal, although no genetic differences were apparent between the southern group and the two source populations (Figure 2). Furthermore, within both the northern and southern groups, hatchery populations showed consistently greater scatter than wild populations, indicating that they may have experienced stronger genetic drift.

We next used a model-based Bayesian clustering approach implemented in fineRADstructure to infer population structure via shared ancestry. The resulting cladogram and coancestry matrix shown in Figure 3 confirmed the results of the AMOVA and PCA while also uncovering the presence of more subtle structuring. Specifically, two major clades were identified. The first of these, shown on the left of the cladogram and represented by a cluster of individuals in the bottom left of the coancestry matrix, comprised individuals from the northern group. This was further sub-divided into two distinct clusters, the first comprising mainly individuals from the Seasalter, Maldon and Bangor hatcheries in the UK and the second comprising mostly wild individuals from Germany and Scandinavia. The remaining individuals were grouped together into a second major clade shown on the right of the

cladogram, which in turn was sub-divided into three main clusters comprising the southern group and source populations, oysters from the Guernsey, Seasalter and two of the French hatcheries, and Portugal. The fact that samples from Seasalter were distributed across two different clusters is consistent with the fact that oysters have been exchanged between Seasalter and Guernsey (Mark Dravers, pers. comm.). Most of the individuals from the remaining French hatchery populations could also be clearly distinguished within the southern group. Samples from Scotland were in different part of the cladogram, but clustered always with other samples from hatcheries. Furthermore, levels of shared coancestry varied appreciably across the dataset, with oysters from the Guernsey hatchery having the highest levels, the remaining hatchery populations as well as oysters from Portugal having intermediate levels, and wild individuals having the lowest levels.

Variation in inbreeding

To explore patterns of inbreeding among wild and hatchery populations, we calculated genomic inbreeding coefficients for all of the individuals in our dataset.. Mean genomic inbreeding coefficients were strongly positively correlated with average pairwise relatedness values within populations (linear regression, $b = 1.39$, $r^2 = 0.79$, $p < 0.001$, Figure 4), which in turn were tightly correlated with mean shared coancestry values extracted from the fineRADstructure analysis (linear regression, $b = 48.99$, $r^2 = 0.99$, $p < 0.001$). This indicates that, as expected, genomic inbreeding coefficients tend to be higher in populations with elevated levels of relatedness and shared coancestry.

Pooling individuals according to the six main groups described in the first paragraph of the results revealed highly significant differences (Figure 5a, Kruskal-Wallis test: $\chi^2 = 105.17$, d.f. = 4, $p < 0.001$). First of all, regardless of whether wild or hatchery populations were considered, a significant tendency was found for genomic inbreeding coefficients to be higher in the northern than in the southern group (*post hoc* pairwise Mann-Whitney tests: wild: adjusted $p < 0.001$, hatchery: adjusted $p = 0.005$). This is consistent with previous studies showing that genetic diversity is lower in the northern group (see discussion). Second, genomic inbreeding coefficients were significantly higher in hatchery versus wild populations (*post hoc* pairwise Mann-Whitney tests: northern group: adjusted p

< 0.001, southern group: adjusted $p < 0.001$). In line with this, identity disequilibrium was higher in individuals sampled from hatchery ($g_2 = 0.0065$, bootstrap 95% confidence interval = 0.0044–0.0089) than from wild populations ($g_2 = 0.0022$, bootstrap 95% confidence interval = 0.0014–0.0029), indicating that hatchery reared oysters have greater variance in inbreeding (Figure 5b). This reflects a general tendency for the variance in inbreeding to be higher both within and among hatcheries relative to wild populations (Figure 5c). The highest genomic inbreeding coefficients were found in oysters from Guernsey, while in France and England, hatcheries with both intermediate (e.g. Fh1, FH3, SES and MAL) and low (e.g. IFR, FH2, FH4, WAL and SCO) levels of inbreeding were present.

RAD sequencing

Although we did not expect our results to be strongly affected by ascertainment bias because oysters from both the northern and southern groups were used in the discovery panel for the SNP array (Gutierrez et al. 2017), we nevertheless generated for comparison a parallel genetic dataset comprising RAD sequencing data for 40 individuals from eight populations (see Materials and methods for details). This resulted in a total of 869,113,776 high quality paired-end reads that were assembled into 697,354 RAD loci. From these, we called a total of 7,322,935 SNPs, of which 115,087 were retained for further analyses after filtering. We found a virtually identical pattern of population structure, with the PCA clearly discriminating the northern group from the southern group, the wild from the hatchery populations, and oysters from Portugal (Figure 6a and b). Similarly, fineRADstructure identified four main groups comprising the southern group and source population, the northern group, Portugal and the Scottish hatchery population (Figure 6c). Genomic inbreeding coefficients based on the RAD data showed a similar pattern to those calculated from the SNP array (Figure 6d), although inbreeding levels appeared to be somewhat higher for the Danish and Norwegian populations. Notably, oysters from Portugal had the highest levels of inbreeding in our RAD analysis, which would not be expected if our sample contained hybrids. Nevertheless, due to the high pairwise F_{ST} values obtained in all comparisons involving oysters from Portugal (Supplementary Table 3), we cannot exclude that these samples may actually be pure *C. angulata*, rather than pure *C. gigas*.

Discussion

We used a medium density SNP array to characterize the genetic structure of Pacific oyster populations across Europe as well as to evaluate levels of inbreeding in wild and hatchery populations. Our comprehensive sampling design coupled with high-resolution genomic data allowed us to resolve patterns of genetic differentiation over both broad and fine geographic scales. Specifically, we found clear support for a northern and southern European group, with the latter being virtually identical to the Japanese and Canadian source populations, in support of previous studies (Huvet et al. 2000, Moehler et al. 2011, Rohfritsch et al. 2013). We furthermore resolved substantial genetic differences between wild and hatchery populations and compared genomic inbreeding coefficients to show that certain hatcheries have higher levels of inbreeding than in the other populations. Given that *C. gigas* carries a high genetic load that has been proposed to be responsible for substantial early mortality (Launey & Hedgecock 2001, Taris et al. 2007, Plough & Hedgecock 2011), we believe that our findings could have important implications for aquaculture.

Population structure

Several studies have investigated the population structure of Pacific oysters in Europe and interpreted their findings in the light of the known and rather complex history of multiple introductions and invasions. Our research compliments and builds upon these studies in a number of ways. First, we were able to confirm previous findings based on mitochondrial DNA as well as small panels of nuclear markers (Huvet et al. 2000, Moehler et al. 2011, Meisterheim et al. 2013, Rohfritsch et al. 2013, Lallias et al. 2015) that European Pacific oyster populations are broadly structured into northern and southern groups. Although this is not necessarily surprising, studies based on one or a few markers can suffer from biases related to stochastic processes (Rokas & Carroll 2005). Consequently, our study lends further weight to the conclusion that the north-south divide is a genome-wide phenomenon that is robust to different methodologies and repeatable across studies.

We were also able to confirm previous studies (Huvet et al. 2000, Moehler et al. 2011, Rohfritsch et al. 2013) reporting negligible genetic differentiation between the source population of Japan and the southern European group. Despite having analyzed samples from both Japan and British Columbia, which was a secondary site of introduction into Europe (Wolff & Reise 2002), and having several orders of magnitude higher genetic resolution than previous studies, both PCA and fineRADstructure failed to find any clear differences between the southern group and source populations. Furthermore, although a number of comparisons involving Japan and British Columbia yielded significant F_{st} values, the magnitude of these estimates was low. Our results therefore lend additional weight to the notion that Pacific oysters did not experience a pronounced founder effect when they were introduced into southern Europe. This is consistent with the observation that many thousands tonnes of spat were transferred into northern France from Japan as well as many hundreds of tonnes of adults from British Columbia (Grizel & Héral 1991, Nehring 1999, Wolff & Reise 2002).

In addition to confirming previous findings, our genomic data also allowed us to resolve fine-scale patterns that could not be detected in previous studies. In particular, Huvet et al. (2004), Meisterheim et al. (2013) and Rohfritsch et al. (2013) did not find any significant genetic differences among populations of the southern group, regardless of whether mitochondrial or nuclear markers were used. This apparent homogeneity was attributed to the prodigious reproductive potential of this species coupled with the possession of long-lived pelagic larvae and frequent transfers of farmed stocks (Meisterheim et al. 2013, Rohfritsch et al. 2013). By contrast, we not only found that a number of pairwise population comparisons within the southern group yielded significant F_{st} values (Supplementary Table 3) but also uncovered a significant isolation-by-distance pattern among the wild populations from southern Europe. The highest F_{ST} values in Southern Europe were obtained for comparisons involving Italy, presumably due to the fact that *C. gigas* was introduced into this part of the Mediterranean during the late 1960s (Šegvić-Bubić et al. 2016). We also found some evidence for the presence of fine-scale population structure within the Northern cluster, although this was more equivocal. Specifically, most but not all of the individuals from Scandinavia and Germany clustered apart from the British hatcheries in the fineRADstructure analysis, although this distinction was not

readily apparent in the PCA. Such a pattern is consistent with Pacific oysters having been imported repeatedly from UK hatcheries to Germany and Scandinavia (d'Auriac et al. 2017).

Comparison of wild and hatchery populations

Two major innovations of our study were first to sample wild and hatchery populations extensively enough to facilitate a meaningful and broad-scale comparison, and second to quantify inbreeding directly from genomic data. Repeated introductions of genetic material from different aquaculture broodstocks are commonplace and should in principle contribute towards the genetic homogenization of wild populations and hatcheries (Moehler et al. 2011). Moreover, a certain degree of genetic exchange between wild and hatchery populations can be expected, at least in France where oyster production in some hatcheries is partially based on wild-caught spat and natural reproduction of farmed oysters occurs (Pouvreau et al. 2016). Set against this, however, temporal sweepstake effects (Hedgecock & Pudovkin 2011) and far smaller numbers of breeding individuals in aquaculture populations (Kochmann et al. 2012) could potentially increase genetic drift and drive genetic differentiation from wild populations. Our data lend support to the latter scenario as we found that hatcheries showed a clear tendency to cluster apart from wild populations and were also characterized by elevated levels of shared coancestry and inbreeding.

Although small panels of genetic markers like microsatellites are capable of resolving population structure, under most circumstances they provide poor estimates of inbreeding (Balloux et al 2004). This has greatly hindered the study of inbreeding in wild populations lacking pedigrees (Pemberton 2008). Consequently, we used our SNP data to calculate genomic inbreeding coefficients for the first time to our knowledge for a marine invertebrate in order to investigate how aquaculture practices may have influenced levels of inbreeding in oyster hatcheries. We uncovered a clear tendency for both the magnitude of inbreeding and its variance to be higher in hatchery populations. This might be considered surprising given that *C. gigas* is produced in vast numbers and is capable of long-distance dispersal mediated by free-swimming planktotrophic larvae (FAO, http://www.fao.org/fishery/culturedspecies/Crassostrea_gigas). However, Pacific oysters also have

one of the smallest documented effective to census population size ratios (10^{-6} , Frankham 1997) reflecting a general tendency in marine invertebrates for highly variable sweepstakes reproductive success resulting from a combination of high fecundity and low larval survivorship (Hedgecock & Pudovkin, 2011). Concretely, a single oyster can produce several tens of millions eggs in a single season (FAO, http://www.fao.org/fishery/culturedspecies/Crassostrea_gigas), but mortality rates within commercial oyster hatchery cultures can be as high as 98% between fertilization and the spat stage (Plough & Hedgecock 2011), leading to high variance in reproductive success of hatchery broodstock (Boudry et al. 2002)

Our findings are in line with a previous study documenting lower microsatellite allelic diversity in hatchery-reared relative to wild individuals within Loch Foyle in Northern Ireland (Kochmann et al. 2012), although a similar study did not find any differences between wild and aquaculture populations in the Wadden Sea (Moehler et al 2011). Moreover, heterozygote deficiency has been observed in several previous studies of oysters (e.g. Lallias et al. 2010, Meisterheim et al. 2013, Rohfritsch et al. 2013, Lallias et al. 2015), which has been interpreted as being suggestive of inbreeding (Faust et al. 2017). Finally, experimental studies have observed massive distortions in marker segregation ratios in F_2 oyster families, consistent with a high genetic load comprising multiple recessive mutations under strong viability selection (Launey & Hedgecock 2001, Plough & Hedgecock 2011). Hence, our study contributes towards a growing body of evidence in support of Launey & Hedgecock's (2001) argument that inbreeding may be a biologically and economically important phenomenon in oysters as well as possibly in other marine invertebrates.

It is important to recognize that not all of the hatchery reared oysters in our study showed higher levels of inbreeding than wild populations. By implication, inbreeding is not associated with hatchery propagation *per se* but may rather arise due to differences in management practices among facilities, which in many cases will reflect differing priorities. For example, many hatcheries minimize the risk of inbreeding by enhancing their broodstock with oysters collected from the wild (Emmanuel Vernier, pers. comm.) or by maximizing numbers of effective breeders, while others actively avoid these

practice (Mathieu Montergous, pers. comm.), presumably to minimize the risk of disease transmission. In other cases, it may be desirable to maintain particular families or lineages that have been selected based on specific characteristics, even if this results in somewhat higher levels of consanguinity (Mark Dravers, pers. comm.).

Practical implications for oyster aquaculture

Inbreeding is known to negatively impact a multitude of commercially relevant fitness traits, from individual growth rate through harvest body size to survival, in many aquaculture organisms (Gallardo et al. 2004, Deng et al. 2005, Moss et al. 2007, Lyu et al. 2018). More specifically, previous studies of oysters have found strong inbreeding depression for early viability (Plough & Hedgecock 2011) as well as yield, growth rate and survival to harvest in adults (Evans et al. 2004). Consequently, elevated inbreeding levels in certain hatcheries are worthy of further exploration and it may be worth considering intervention strategies aimed at increasing genetic diversity.

With respect to the need for further exploration, it is worth bearing in mind that although our total sample size of oysters was reasonably large given the number of markers deployed, only around ten samples were analyzed on average from each population. While this is unlikely to have appreciably effected our inference of population structure (Willing et al. 2012), the inference of inbreeding levels within populations could be sensitive to the inadvertent sampling of highly related individuals when sample sizes are small. Furthermore, high variance in reproductive success within and across generations could potentially lead to different cohorts from the same hatcheries varying substantially in their levels of inbreeding. Consequently, it would be worthwhile enlarging sample sizes within hatcheries, as well as collecting and analyzing samples from the same locations over multiple years in order to provide more robust inferences of average inbreeding levels and allow these to be interpreted in the light of temporal variation. Such a study would further benefit from the development of a larger SNP array and a more contiguous *C. gigas* genome assembly, which would allow inbreeding to be evaluated with even greater precision through the use of mapped genetic markers to quantify runs of homozygosity.

Having done so, a useful next step would be to evaluate in greater detail the effects of inbreeding on commercially important traits within hatcheries. So far, only a single study has evaluated the effects of inbreeding on adult traits in a commercial growing environment and this was based on a crossing design that maximised variation in inbreeding among families (Evans et al. 2004). By contrast, our approach of quantifying inbreeding directly from genomic data could in principle circumvent the need for experimental crosses, therefore allowing inbreeding depression to be directly quantified in real hatchery populations when phenotypic data are available. This could help to inform hatchery managers about the potential costs of inbreeding and the possible benefits of intervention strategies.

Finally, a number of potential intervention strategies aimed at reducing inbreeding and increasing genetic diversity could be envisaged. The first obvious approach would be to incorporate individuals from wild populations into hatchery broodstocks, as also discussed by Lallias et al. (2010) in the context of flat oysters. However, caution is warranted as selective breeding in captivity may lead to adaptive changes that are absent from wild populations (Lachambre et al. 2017) so the fitness consequences of such crosses remain unclear. A second possibility would be exchange individuals more extensively among hatcheries. Within Europe, the practice of exchanging oyster stocks between different countries is becoming more common, but we are not aware of any such exchanges between the United Kingdom and the European mainland, probably due to the perceived risk of disease transmission. Clearly, hatchery managers need to balance the pros and cons of selective breeding and maximising genetic diversity, but either way genomic tools such as high density SNP arrays provide a means of evaluating the genetic consequences of chosen management practices.

Caveats

SNP arrays provide a cost-effective and convenient route to genome-wide investigations but can be prone to ascertainment bias when the samples used in the initial SNP discovery phase differ from those being interrogated on the array (Lachance & Tishkoff 2013). However, we believe this is unlikely to substantially affect our main conclusions for two reasons. First, the discovery panel of individuals used to construct the array was unusually large, comprising over 200 individuals from

eight different localities. By implication, much of the genetic diversity of the species in Europe should have been captured, including rare alleles that may easily be missed with smaller discovery panels but which could potentially be present at higher frequencies in unsampled populations. Second, although the discovery panel comprised primarily individuals from hatcheries, the northern and southern groups were roughly equally represented and we therefore have no reason to expect any broad-scale biases to be present. Two further points should also be recognized. First, our analyses of population structure will if anything be conservative, as ascertainment bias should lead to the underestimation of genetic differentiation when peripheral populations carry previously undetected alleles. Second, ascertainment bias cannot explain higher levels of inbreeding in hatchery populations nor variation in inbreeding among hatcheries in the UK and France. This is because all of these populations were used to generate the array, so ascertainment bias if present would be expected to generate the opposite pattern of increased homozygosity in wild populations.

Nevertheless, we took into account the conservative possibility that ascertainment bias could be responsible for the ostensibly high level of inbreeding in the putatively hybrid Portuguese population. To test this possibility as well as to confirm our broader findings, we RAD sequenced a subset of individuals and repeated all of our analyses. Our previous results were largely confirmed, with virtually identical patterns of population structure and inbreeding being obtained, lending further weight to our main conclusions. Furthermore, oysters from Portugal were again found to have relatively high genomic inbreeding coefficients based on the RAD data. As we would expect hybrids to be relatively outbred, this finding points towards hybridisation between *C. gigas* and *C. angulata* being negligible in our sample. Consequently, it appears that the Portuguese oysters could represent an inbred and isolated *C. gigas* population. However, we cannot discount the further possibility that we inadvertently sampled *C. angulata* from this location, as the two species are morphologically indistinguishable, the SNP array may include loci that cross amplify in *C. angulata* (Gagnaire et al. 2018), and F_{st} comparisons involving our Portuguese sample were consistently high.

Conclusion

We harnessed some of the latest developments in genomics to shed new light on the population structure of Pacific oysters along a European latitudinal cline as well as to compare levels of inbreeding between wild and hatchery populations. The several orders of magnitude higher genetic resolution provided by the medium density SNP array allowed us not only to confirm previous findings (Meistertzheim et al. 2013, Rohfritsch et al. 2013, Lallias et al. 2015, Faust et al. 2017) but also to detect fine-scale patterns including genetic differences between wild and hatchery populations. We furthermore uncovered evidence that some hatchery practices may exacerbate inbreeding, which could have important economic consequences and should be managed. Finally, our study contributes to a growing consensus that inbreeding could be more prevalent in animal populations than previously envisaged (Keller & Waller 2002), even in highly fecund species with high dispersal.

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Data archiving statement

Individual genotypes files obtained from the SNP array and from the analyses of the RAD sequencing data are available from the Dryad Digital Repository: <https://doi.org/10.5061/dryad.6d778b6>

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Figures legends

Figure 1: Map showing Pacific oyster sampling locations. The two source populations from the Miyagi Prefecture in Japan and British Columbia in Canada are indicated with yellow circles represent. Wild and hatchery populations within the southern group are indicated in red and orange respectively, while wild and hatchery populations within the northern group are represented by blue and purple circles respectively. Finally, the population from Portugal, where hybridization between *C. gigas* and *C. angulata* is known to occur, is indicated by a green circle.

Figure 2: Scatterplot showing individual variation in principal component (PC) scores derived from a principal component analysis (PCA) of the genomic data. Panel (a) shows PC1 plotted against PC2, and panel (b) shows PC1 plotted against PC3. The amounts of variation explained by each PC are given as percentages. Samples are color coded as described in the legend of Figure 1.

Figure 3: Output of the fineRADstructure analysis of the genomic data. In the cladogram, branches are color coded according to sampling origin as shown in Figure 1 and nodes with greater than 95% bootstrap support are marked by black points. The heatmap indicates pairwise coancestry between individuals, with blue and purple representing the highest levels, red and orange indicating intermediate levels and yellow representing the lowest levels of shared coancestry.

Figure 4: Relationship between average pairwise relatedness and average genomic inbreeding coefficients within populations (linear regression, $b = 1.39$, $r^2 = 0.79$, $p < 0.001$).

Figure 5: Levels of inbreeding in wild and farmed populations inferred from genome-wide SNP data. Panel (a) shows differences between wild and hatchery samples from the northern and southern groups separately as well as for the source populations. Raw data points are shown together with standard Tukey box plots. Panel (b) shows bootstrapped g_2 values for individuals sampled from wild (dark grey) versus hatchery (light grey) populations. The empirical g_2 values and their corresponding 95% confidence intervals are depicted by dashed vertical lines and horizontal bars respectively. Panel (c) shows population-specific variation in inbreeding. In panels (a) and (c), the populations are color coded as described in the legend of Figure 1.

Figure 6: Results of repeated analyses of population structure and inbreeding based on a subset of RAD sequenced individuals from eight populations (see Methods for details). Panels (a) and (b) show individual variation in principal component (PC) scores derived from a principal component analysis. Panel (a) shows PC1 plotted against PC2 while panel (b) shows PC1 plotted against PC3. The amounts of variation explained by each PC are given as percentages. Panel (c) presents the output of

the fineRADstructure analysis, including both the cladogram and the heatmap representing pairwise coancestry values between individuals. Panel (d) shows population-specific variation in inbreeding. Samples are color coded as described in the legend of Figure 1.

Supplementary materials

Supplementary figure 1: Change in the number of polymorphic *de novo* assembled loci present in at least 80% of the samples for increasing values of the $-M$ parameter. The red and the blue lines correspond to assemblies performed by setting $-n = -M$ and $-n = -M + 1$ respectively. At $-M = 5$, the number of assembled loci reached a plateau and was therefore selected for the analysis of the full dataset. $-n = 6$, was chosen over $-n = 5$ because it yielded a higher number of loci.

Supplementary Table 1: List of thresholds for quality assessment recommended by Affimetrix best practice workflow.

Supplementary Table 2: Table of populations selected for RAD sequencing, including coordinates, origin classified as either wild or hatchery, and the number of samples that were retained for analysis after quality control and filtering.

Supplementary Table 3: Table showing pairwise F_{ST} values (below diagonal) and respective p -values (above diagonal) for pairwise population comparisons. Values indicated in bold are significant after table-wide Bonferroni correction for multiple tests.

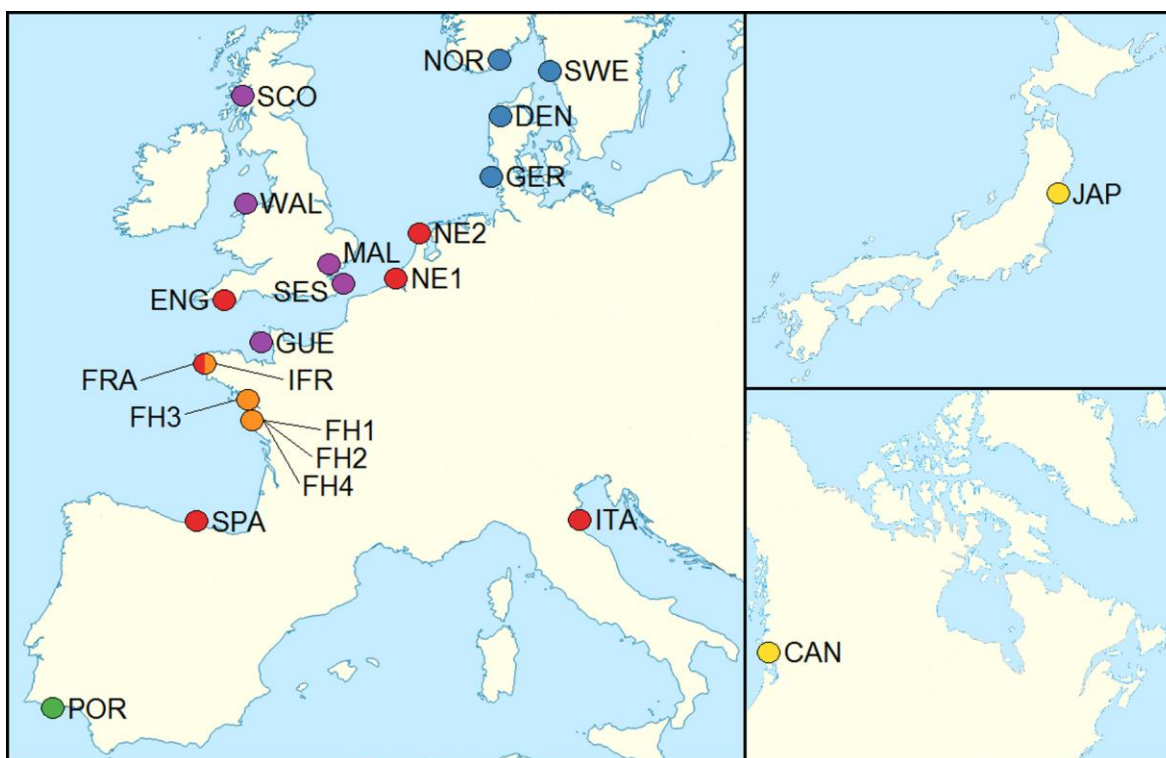
Table 1: Table of sampling locations including coordinates, origin classified as either wild or farmed, and the number of samples that were retained for analysis after quality control.

Population ID	Location	Latitude	Longitude	Origin	Sample size passing QC
POR	Faro, Portugal	37.002	-7.583	Wild	9
ITA	Ravenna, Italy	44.402	12.122	Wild	10
SPA	Santoña, Spain	43.426	-3.543	Wild	8
FRA	Brest, France	48.215	-4.462	Wild	10
ENG	Plymouth, UK	50.373	-3.441	Wild	9
NE1	Oosterschelde, Netherlands	51.608	3.91	Wild	12
NE2	Texel, Netherlands	53.001	4.474	Wild	10
IFR	Ifremer, France	48.351	-4.551	Hatchery	12
FH2	SATMAR, France	46.948	-2.052	Hatchery	10
FH4	Novostrea, France	46.954	-2.044	Hatchery	10
FH1	Marinove, France	46.987	-2.238	Hatchery	10
FH3	France Naissain, France	47.514	-2.666	Hatchery	10
GUE	Guernsey, UK	49.497	-2.502	Hatchery	10
SCO	Oban, UK	55.534	-5.244	Hatchery	8
SES	Sea Salter, UK	51.378	1.212	Hatchery	9
MAL	Maldon, UK	51.724	0.71	Hatchery	9
WAL	Bangor, UK	53.098	-4.15	Hatchery	6
GER	Sylt, Germany	55.152	8.253	Wild	12
DEN	Limfjorden, Denmark	56.833	8.906	Wild	11
SWE	Kristineberg, Sweden	58.134	58.134	Wild	12
NOR	Arendal, Norway	58.428	8.793	Wild	11
JAP	Matsushima, Japan	38.367	141.066	Wild	12
CAN	Vancouver, Canada	50.164	-124.432	Wild	12
All	-	-	-	-	232

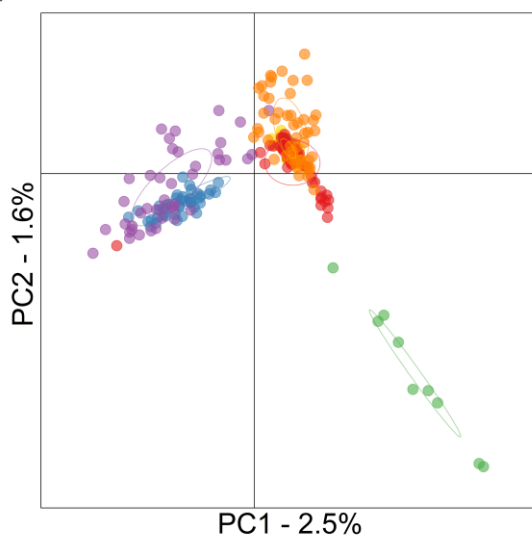
Table 2: Results of the hierarchical analysis of molecular variance (AMOVA).

Source of variation	d.f.	Sum of squares	% variation	Φ	p-value
Among regions	3	48,727.66	1.7	0.017	0.024
Between origins within regions	2	17,734.93	0.39	0.004	0.005
Among populations within origins	17	116,998.80	2	0.02	0.001
Among samples within populations	209	1,014,805.34	0.22	0.002	0.406
Within samples	232	1,121,225.26	95.69	0.043	0.001
Total	463	2,319,491.99	100	-	-

Five different hierarchical levels were evaluated. First, the dataset was divided into four 'regions' corresponding to the southern group, the northern group, the source populations and Portugal. Next, each region was divided into 'origins' depending whether the samples were from wild or farmed populations. Finally, the remaining variance was partitioned among sampling locations, individuals within sampling locations, and within individuals.



a)



b)

